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the biosynthetic pathway of the proinflammatory prostaglandins and a factor that has been implicated in seizure

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Donald Kuhn 19b. TELEPHONE NUMBER 313-576-4457

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Report Title

Microglia as primary mediators of nerve agent neuropathy

ABSTRACT

Nerve agent-induced seizures cause neuronal damage in brain limbic and cortical circuits leading to persistent behavioral and cognitive deficits. Without aggressive anticholinergic and benzodiazepine therapy, seizures can be prolonged and neuronal damage progresses for extended periods of time. The objective of this study was to determine the effects of the nerve agent soman on expression of cyclooxygenase-2 (COX-2), the initial enzyme in the biosynthetic pathway of the proinflammatory prostaglandins and a factor that has been implicated in seizure initiation and propagation. Rats were exposed to a toxic dose of soman and scored behaviorally for seizure intensity. Expression of COX-2 was determined throughout brain from 4 hr to 7 days after exposure by immunohistochemistry and immunoblotting. Microglial activation and astrogliosis were assessed microscopically over the same time-course. Soman increased COX-2 expression in brain regions known to be damaged by nerve agents (e.g., hippocampus, amygdala, piriform cortex and thalamus). COX-2 expression was induced in neurons, and not in microglia or astrocytes, and remained elevated through 7 days. The magnitude of COX-2 induction was correlated with seizure intensity. COX-1 expression was not changed by soman. Increased expression of neuronal COX-2 by soman is a late-developing response relative to other signs of acute physiological distress caused by nerve agents. COX-2-mediated production of prostaglandins is a consequence of the seizure-induced neuronal damage, even after survival of the initial cholinergic crisis is assured. COX-2 inhibitors should be considered as adjunct therapy in nerve agent poisoning to minimize nerve agent-induced seizure activity.

List of papers submitted or published that acknowledge ARO support during this reporting period. List the papers, including journal references, in the following categories:

(a) Papers published in peer-reviewed journals (N/A for none)

Angoa-Perez, M., Kreipke, C.W., Thomas, D.M., Van Shura, K.E., Lyman, M., McDonough, J.H. and Kuhn, D.M. Soman increases neuronal COX-2 levels: Possible link between seizures and protracted neuronal damage. Neurotoxicology, in press, 2010. PMID: 20600289.

Number of Papers published in peer-reviewed journals: 1.00

1.00

(b) Papers published in non-peer-reviewed journals or in conference proceedings (N/A for none)

Number of Papers published in non peer-reviewed journals: 0.00

(c) Presentations

Angoa-Perez, M., Verbeem, D.M., Thomas, D.M., Van Shura, K., Lyman, J.H., McDonough, J.H., and Kuhn, D.M. The nerve agent sarin causes widespread microglial activation in brain. Soc. Neurosci., 154.7, 2008.

Number of Presentations:

Non Peer-Reviewed Conference Proceeding publications (other than abstracts):

Number of Non Peer-Reviewed Conference Proceeding publications (other than abstracts):

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(d) Manuscripts

Number of Manuscripts: 0.00

Patents Submitted

Patents Awarded

Graduate Students

NAME PERCENT SUPPORTED

FTE Equivalent: Total Number:

Names of Post Doctorates

<u>NAME</u>	PERCENT_SUPPORTED
Mariana Angoa-Perez	0.25
FTE Equivalent:	0.25
Total Number:	1

Names of Faculty Supported

NAME	PERCENT SUPPORTED	National Academy Member
Donald M. Kuhn	0.10	No
FTE Equivalent:	0.10	
Total Number:	1	

Names of Under Graduate students supported

NAME PERCENT_SUPPORTED

FTE Equivalent: Total Number:

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This section only applies to graduating undergraduates supported by this agreement in this reporting period

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Statement of the problem studied:

Nerve agents can cause seizures after acute intoxication and these seizures can lead eventually to neuronal damage. The purpose of the present study was to assess the extent to which seizure development after exposure of rats to soman resulted in the up-regulation of cyclooxygenase 2 (COX-2) in brain areas known to be damaged by soman.

Summary of the results:

Soman causes a time-dependent increase in COX-2 expression

The effects of soman on COX-2 protein expression were examined by immunohistochemical analysis over a broad time course (i.e., 4 hr to 7 days after soman). Counts of COX-2 positive cells revealed a somewhat delayed response to soman (Fig 1). Very few cells expressing COX-2 were seen from 4-12 hr after soman treatment (data not shown). By 24-48 hr, large numbers of COX-2 positive cells were seen in hippocampal CA3 regions and especially the dentate gyrus (Fig. 1A and 1B) as well as in the amygdala (Fig. 1C) and piriform cortex (Fig. 1D). By 7 days COX-2 levels declined slightly below those seen at 48 hr in each brain region, but remained significantly elevated over control. Increases in COX-2 immunoreactivity were observed in cingulate cortex and ventral thalamus as well (data not shown).

Soman increases COX-2 expression at the cellular level in a highly circumscribed manner as revealed by immunohistochemistry

Immunohistochemical analyses revealed the highly circumscribed effect of soman on COX-2 expression at the cellular level. After treatment with soman (48 hr), COX-2 positive cells essentially define the anatomical facets of the dentate gyrus, CA3 and CA1 regions of the hippocampus (Fig 2). Cells expressing COX-2 immunoreactivity were small and uniformly round. The piriform cortex and amygdala also showed substantial increases in the number of COX-2 immunoreactive cells after soman (Fig 3). These cells were somewhat more diffuse in the piriform cortex and many displayed an extensive axonal network. COX-2 positive cells were more densely packed in the amygdala of soman-treated animals by comparison to the piriform cortex (see Fig 3). Soman did not change the expression of COX-1 at any time (4 hr to 7 days) in any brain region examined (Fig 4). COX-1 immunoreactivity was very weak in hippocampus of controls (Fig. 4A) and soman treated rats (Fig. 4B). COX-1 containing cells were also seen throughout the amygdala with no apparent alteration by soman in their number or in the intensity of their staining for COX-1 (Fig. 4C and 4D).

Soman-induced increases COX-2 protein levels are correlated with seizure intensity

Immunoblot analysis provided independent confirmation of soman effects on COX-2 expression. Soman caused increases in hippocampal COX-2 that varied considerably (Fig 5A). Because all rats were injected with the same soman dose (i.e., 1.2 X LD₅₀), these results suggest that the changes in COX-2 were not linked to soman per se. All rats were scored for seizures as described in Materials and Methods and behavioral scores were plotted versus the fold-increase in COX-2 immunoreactivity on western blots. This analysis indicated that COX-2

expression was positively correlated with seizure intensity (Fig 5B). Soman-treated rats showing no fasciculations, tremors or seizure activity (behavioral score of 0) showed slight increases in COX-2 (~1.5-2 fold over controls). Animals showing mild fasciculations (behavioral score of 1) and tremor (behavioral score of 2) showed increases in COX-2 expression that increased by 4-10 fold. By far, the largest increase in COX-2 expression (7-15-fold) was seen in rats showing the most intense seizures (behavioral score of 3). Similar results for COX-2 expression were seen in other brain regions (data not shown). Immunoblot analyses also confirmed that hippocampal COX-1 protein levels were not altered by soman at any seizure intensity (see Figs 5A and 5B).

Soman increases COX-2 expression in neurons and not in microglia or astrocytes

COX-2 can be expressed in neurons and by activated microglia and astrocytes (Minghetti and Levi, 1998) so efforts were made to identify the cell-type in which COX-2 expression was increased by soman. First, hippocampus was examined 48 hr after soman exposure for changes in astrocyte and microglial reactivity using GFAP and Isolectin B₄. respectively. The density and staining intensity of astrocytes were increased substantially in hippocampus after soman (Fig. 6B) by comparison to controls (Fig. 6A). Microglial activation in hippocampus was also increased dramatically by soman (Fig. 6D) by comparison to controls (Fig. 6C). In light of this soman-induced gliosis in hippocampus, brain sections were labeled with COX-2 antibodies followed by co-labeling with antibodies against either NeuN to identify neurons, antibodies against GFAP to identify astrocytes, or ILB₄ to identify activated microglia. Patterns of COX-2 (Fig. 7A) and NeuN (Fig. 7B) fluorescence in hippocampus were very similar and when merged, a near-total overlap of cells that are immuno-positive for both COX-2 and NeuN was evident (Fig 7C). The soman-induced microglial activation is evident throughout the hippocampus (Fig. 7E) and it is clear from the merged image (Fig 7F) that the pattern of COX-2 fluorescence staining shows no overlap with that of microglia. Finally, intense GFAP reactivity is evident after soman treatment in the area between CA3 and the dentate gyrus of the hippocampus (Fig 7H), and the lack of overlap of COX-2 containing cells with astrocytes is apparent in the merged image (Fig 7I). The morphology of COX-2 containing cells in the hippocampus of soman treated rats also gives a clear indication of their identity as neurons.

Bibliography

Baille V, Clarke PG, Brochier G, Dorandeu F, Verna JM, Four E, Lallement G and Carpentier P. Soman-induced convulsions: the neuropathology revisited. Toxicology 2005; 215:1-24.

Berry WK and Davies DR. The use of carbamates and atropine in the protection of animals against poisoning by 1,2,2-trimethylpropyl methylphosphonofluoridate. Biochem Pharmacol 1970; 19:927-34.

Bezzi P, Carmignoto G, Pasti L, Vesce S, Rossi D, Rizzini BL, Pozzan T and Volterra A. Prostaglandins stimulate calcium-dependent glutamate release in astrocytes. Nature 1998; 391:281-5.

Chapman S, Kadar T and Gilat E. Seizure duration following sarin exposure affects neuro-inflammatory markers in the rat brain. Neurotoxicology 2006; 27:277-83.

Cole-Edwards KK and Bazan NG. Lipid signaling in experimental epilepsy. Neurochem Res 2005; 30:847-53.

Collombet JM, Carpentier P, Baille V, Four E, Bernabe D, Burckhart MF, Masqueliez C, Baubichon D and Lallement G. Neuronal regeneration partially compensates the delayed neuronal cell death observed in the hippocampal CA1 field of soman-poisoned mice. Neurotoxicology 2006; 27:201-9.

Collombet JM, Four E, Bernabe D, Masqueliez C, Burckhart MF, Baille V, Baubichon D and Lallement G. Soman poisoning increases neural progenitor proliferation and induces long-term glial activation in mouse brain. Toxicology 2005; 208:319-34.

Collombet JM, Pierard C, Beracochea D, Coubard S, Burckhart MF, Four E, Masqueliez C, Baubichon D and Lallement G. Long-term consequences of soman poisoning in mice Part 1. Neuropathology and neuronal regeneration in the amygdala. Behav Brain Res 2008; 191:88-94.

Coubard S, Beracochea D, Collombet JM, Philippin JN, Krazem A, Liscia P, Lallement G and Pierard C. Long-term consequences of soman poisoning in mice: part 2. Emotional behavior. Behav Brain Res 2008; 191:95-103.

Dhote F, Peinnequin A, Carpentier P, Baille V, Delacour C, Foquin A, Lallement G and Dorandeu F. Prolonged inflammatory gene response following soman-induced seizures in mice. Toxicology 2007; 238:166-76.

Dillman JF, 3rd, Phillips CS, Kniffin DM, Tompkins CP, Hamilton TA and Kan RK. Gene expression profiling of rat hippocampus following exposure to the acetylcholinesterase inhibitor soman. Chem Res Toxicol 2009; 22:633-8.

Dirnhuber P, French MC, Green DM, Leadbeater L and Stratton JA. The protection of primates against soman poisoning by pretreatment with pyridostigmine. J Pharm Pharmacol 1979; 31:295-9.

Filliat P, Coubard S, Pierard C, Liscia P, Beracochea D, Four E, Baubichon D, Masqueliez C, Lallement G and Collombet JM. Long-term behavioral consequences of soman poisoning in mice. Neurotoxicology 2007; 28:508-19.

Grauer E, Chapman S, Rabinovitz I, Raveh L, Weissman BA, Kadar T and Allon N. Single whole-body exposure to sarin vapor in rats: long-term neuronal and behavioral deficits. Toxicol Appl Pharmacol 2008; 227:265-74.

Kadar T, Shapira S, Cohen G, Sahar R, Alkalay D and Raveh L. Sarin-induced neuropathology in rats. Hum Exp Toxicol 1995; 14:252-9.

Kuehl FA, Jr. and Egan RW. Prostaglandins, arachidonic acid, and inflammation. Science 1980; 210:978-84.

Lee EC. Clinical manifestations of sarin nerve gas exposure. JAMA 2003; 290:659-62.

McDonough JH, Jr., Dochterman LW, Smith CD and Shih TM. Protection against nerve agent-induced neuropathology, but not cardiac pathology, is associated with the anticonvulsant action of drug treatment. Neurotoxicology 1995; 16:123-32.

McDonough JH, Jr., Jaax NK, Crowley RA, Mays MZ and Modrow HE. Atropine and/or diazepam therapy protects against soman-induced neural and cardiac pathology. Fundam Appl Toxicol 1989; 13:256-76.

McDonough JH, Jr., McMonagle J, Copeland T, Zoeffel D and Shih TM. Comparative evaluation of benzodiazepines for control of soman-induced seizures. Arch Toxicol 1999; 73:473-8.

McDonough JH, Jr. and Shih TM. Neuropharmacological mechanisms of nerve agent-induced seizure and neuropathology. Neurosci Biobehav Rev 1997; 21:559-79.

McDonough JH, Jr., Zoeffel LD, McMonagle J, Copeland TL, Smith CD and Shih TM. Anticonvulsant treatment of nerve agent seizures: anticholinergics versus diazepam in soman-intoxicated guinea pigs. Epilepsy Res 2000; 38:1-14.

McDonough JH, Van Shura KE, LaMont JC, McMonagle JD and Shih TM. Comparison of the intramuscular, intranasal or sublingual routes of midazolam administration for the control of soman-induced seizures. Basic Clin Pharmacol Toxicol 2009; 104:27-34.

Minghetti L and Levi G. Microglia as effector cells in brain damage and repair: focus on prostanoids and nitric oxide. Prog Neurobiol 1998; 54:99-125.

Miyaki K, Nishiwaki Y, Maekawa K, Ogawa Y, Asukai N, Yoshimura K, Etoh N, Matsumoto Y, Kikuchi Y, Kumagai N and Omae K. Effects of sarin on the nervous system of subway workers seven years after the Tokyo subway sarin attack. J Occup Health 2005; 47:299-304.

Nishihara I, Minami T, Watanabe Y, Ito S and Hayaishi O. Prostaglandin E2 stimulates glutamate release from synaptosomes of rat spinal cord. Neurosci Lett 1995; 196:57-60.

Ohbu S, Yamashina A, Takasu N, Yamaguchi T, Murai T, Nakano K, Matsui Y, Mikami R, Sakurai K and Hinohara S. Sarin poisoning on Tokyo subway. South Med J 1997; 90:587-93.

Sekiyama N, Mizuta S, Hori A and Kobayashi S. Prostaglandin E2 facilitates excitatory synaptic transmission in the nucleus tractus solitarii of rats. Neurosci Lett 1995; 188:101-4.

Shih T, McDonough JH, Jr. and Koplovitz I. Anticonvulsants for soman-induced seizure activity. J Biomed Sci 1999; 6:86-96.

Shih TM, Duniho SM and McDonough JH. Control of nerve agent-induced seizures is critical for neuroprotection and survival. Toxicol Appl Pharmacol 2003; 188:69-80.

Shih TM and McDonough JH, Jr. Neurochemical mechanisms in soman-induced seizures. J Appl Toxicol 1997; 17:255-64.

Shih TM and McDonough JH, Jr. Organophosphorus nerve agents-induced seizures and efficacy of atropine sulfate as anticonvulsant treatment. Pharmacol Biochem Behav 1999; 64:147-53.

Shih TM, Rowland TC and McDonough JH. Anticonvulsants for nerve agent-induced seizures: The influence of the therapeutic dose of atropine. J Pharmacol Exp Ther 2007; 320:154-61.

Smith WL, DeWitt DL and Garavito RM. Cyclooxygenases: structural, cellular, and molecular biology. Annu Rev Biochem 2000; 69:145-82.

Takemiya T, Maehara M, Matsumura K, Yasuda S, Sugiura H and Yamagata K. Prostaglandin E2 produced by late induced COX-2 stimulates hippocampal neuron loss after seizure in the CA3 region. Neurosci Res 2006; 56:103-10.

Takemiya T, Suzuki K, Sugiura H, Yasuda S, Yamagata K, Kawakami Y and Maru E. Inducible brain COX-2 facilitates the recurrence of hippocampal seizures in mouse rapid kindling. Prostaglandins Other Lipid Mediat 2003; 71:205-16.

Thomas DM, Francescutti-Verbeem DM and Kuhn DM. Methamphetamine-induced neurotoxicity and microglial activation are not mediated by fractalkine receptor signaling. J Neurochem 2008; 106:696-705.

Thomas DM and Kuhn DM. Cyclooxygenase-2 is an obligatory factor in methamphetamine-induced neurotoxicity. J Pharmacol Exp Ther 2005; 313:870-6.

Williams AJ, Berti R, Yao C, Price RA, Velarde LC, Koplovitz I, Schultz SM, Tortella FC and Dave JR. Central neuro-inflammatory gene response following soman exposure in the rat. Neurosci Lett 2003; 349:147-50.

Yamasue H, Abe O, Kasai K, Suga M, Iwanami A, Yamada H, Tochigi M, Ohtani T, Rogers MA, Sasaki T, Aoki S, Kato T and Kato N. Human brain structural change related to acute single exposure to sarin. Ann Neurol 2007; 61:37-46.

Zimmer LA, Ennis M and Shipley MT. Soman-induced seizures rapidly activate astrocytes and microglia in discrete brain regions. J Comp Neurol 1997; 378:482-92.

Fig. 1

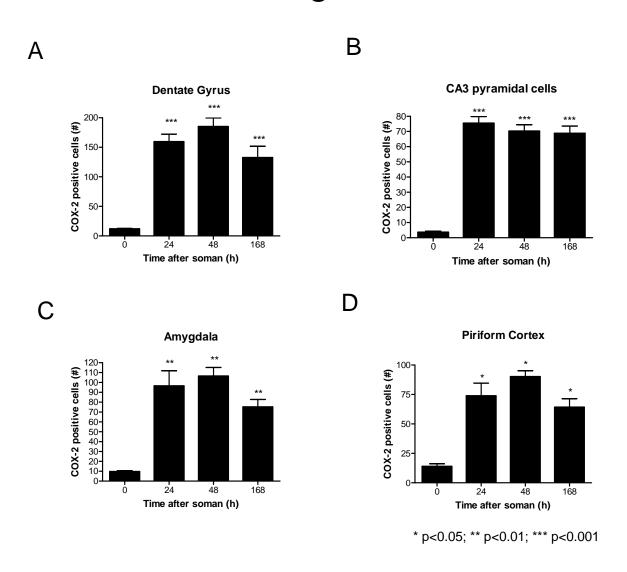


Fig.2

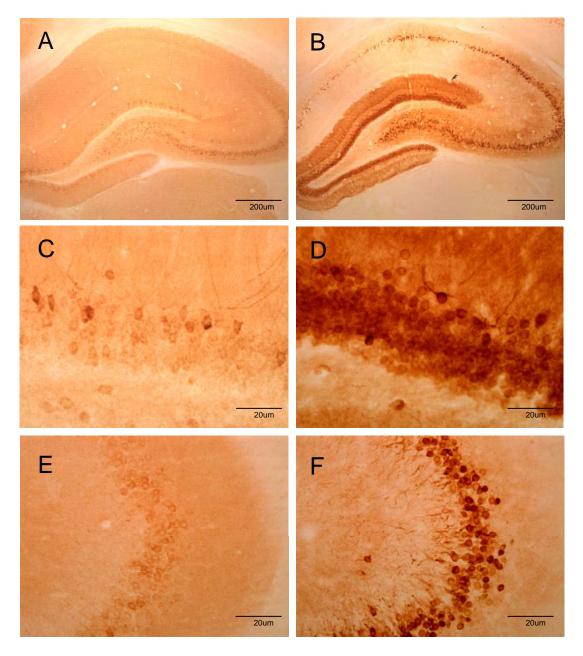


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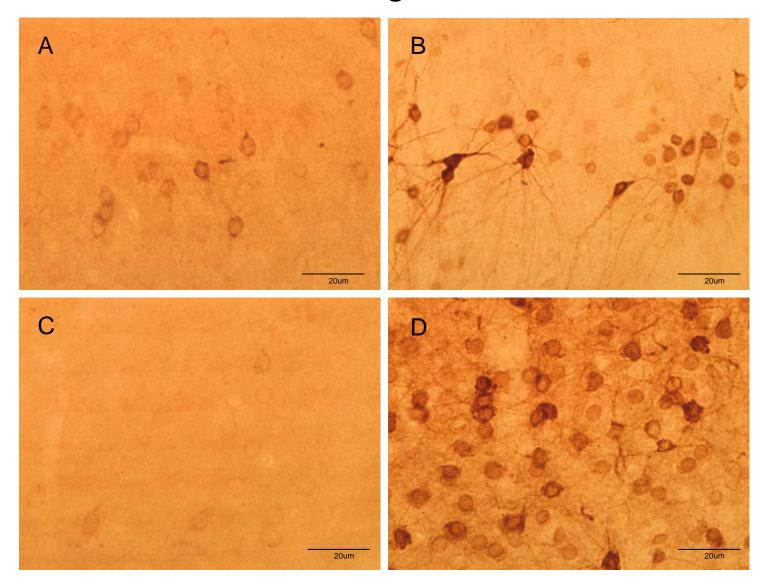


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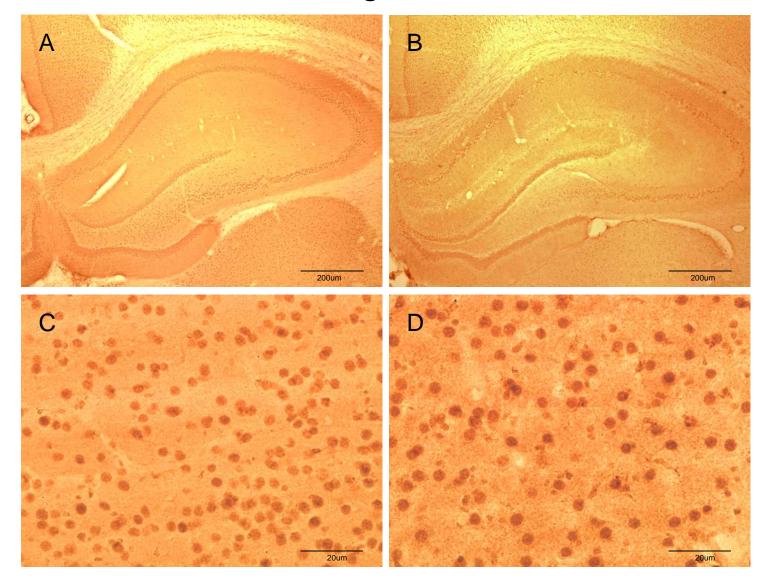


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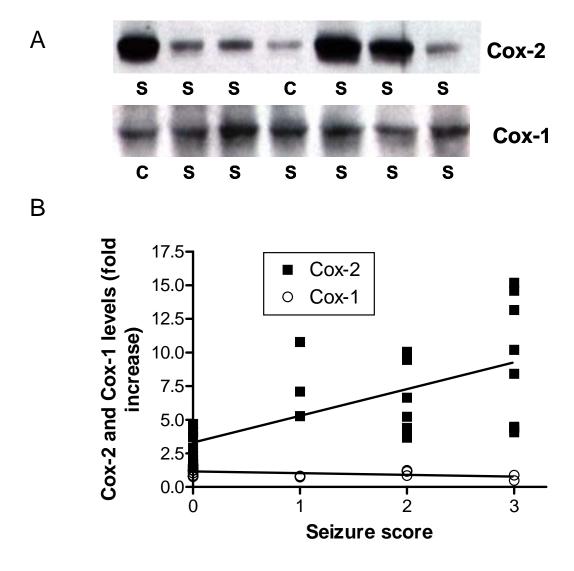


Fig 6.

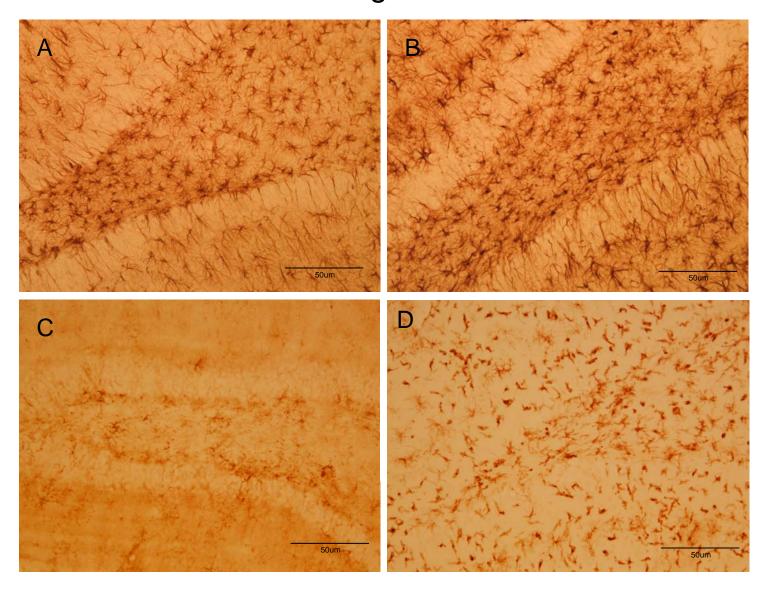


Fig 7.

